

Research

Open Access

## Phylogenetic analysis of Newcastle disease viruses isolated from waterfowl in the Upper Midwest Region of the United States

Naresh Jindal, Yogesh Chander, Ashok K Chockalingam, Martha de Abin, Patrick T Redig and Sagar M Goyal\*

Address: Department of Veterinary Population Medicine, College of Veterinary Medicine, University of Minnesota, 1333 Gortner Avenue, Saint Paul, MN, 55108, USA

Email: Naresh Jindal - jinda014@umn.edu; Yogesh Chander - chand062@umn.edu; Ashok K Chockalingam - chock006@umn.edu; Martha de Abin - fuent006@umn.edu; Patrick T Redig - redig001@maroon.tc.umn.edu; Sagar M Goyal\* - goyal001@umn.edu

\* Corresponding author

Published: 5 November 2009

Received: 14 July 2009

*Virology Journal* 2009, **6**:191 doi:10.1186/1743-422X-6-191

Accepted: 5 November 2009

This article is available from: <http://www.virologyj.com/content/6/1/191>

© 2009 Jindal et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** This study was conducted to characterize Newcastle disease virus (NDV) isolates obtained from waterfowl from the Upper Midwest region of the United States. A total of 43 NDVs were isolated by inoculation of cloacal samples in embryonated chicken eggs. These isolates were obtained from 24 mallards, seven American green-winged teals, six northern pintails, four blue-winged teals, and two wood ducks. Partial sequences of fusion gene were analyzed to determine the pathotypes and genotypes involved.

**Results:** Deduced amino acid sequence of the cleavage site of fusion (F) protein revealed that all isolates had avirulent motifs. Of the 43 isolates, 23 exhibited sequence motif of <sup>111</sup>GGKQGRL<sup>117</sup> at the cleavage site, 19 exhibited <sup>111</sup>GEKQGRL<sup>117</sup> while one isolate showed <sup>111</sup>GERQGRL<sup>117</sup>. Phylogenetic analysis based on comparison with different classes of NDVs revealed that all 43 isolates clustered with class II NDVs and none with class I NDVs. Within class II, five isolates were phylogenetically close to genotype I NDVs while the remaining 38 were close to genotype II.

**Conclusion:** We conclude that more than one genotype of NDV circulates in waterfowl in the Upper Midwest region of the US. Continuous surveillance may help better understand the epidemiology of NDVs maintained in wild bird populations and their relationship to NDVs in domestic poultry, if any.

### Background

Avian paramyxoviruses (APMV) belong to genus *Avulavirus* in the family *Paramyxoviridae*. The genome of APMV is an approximately 15 kb long, negative-sense, single-stranded RNA molecule. It has six genes that encode for a nucleoprotein (N), a phosphoprotein (P), a matrix protein (M), a fusion protein (F), an attachment protein called hemagglutinin-neuraminidase (HN), and a large

polymerase protein (L) [1]. Nine serotypes of avian paramyxoviruses (APMV-1 to APMV-9) have been identified. Of these, APMV-1, also called the Newcastle disease virus (NDV), is the causative agent of Newcastle disease (ND) in poultry. Based on genetic and antigenic analyses of NDV isolates, two major classes (class I and class II) are identified [2,3] and each class has nine genotypes (1-9 genotypes in class I and I-IX in class II) [4,5].

The NDV can cause clinical signs varying from subclinical infections to 100% mortality, depending on the susceptibility of the host and the virulence of the virus. The virus is categorized into velogenic (velogenic neurotropic or velogenic viscerotropic), mesogenic, lentogenic, and asymptomatic enteric strains on the basis of their pathogenesis and virulence. The velogenic strains cause acute fatal infection of chickens of all age groups with clinical findings of nervous signs or extensive hemorrhagic lesions in the gastrointestinal tract. The mesogenic strains are of intermediate virulence and cause moderate respiratory signs with occasional nervous signs while the lentogenic strains cause mild to inapparent infections [1]. The lentogenic strains have been detected in both domestic poultry [6-8] and wild bird populations [4,8,9]. Though velogenic strains are considered exotic (exotic Newcastle disease, END) to US poultry, these strains have been isolated occasionally from different avian species in the US [10,11]. During 2002-2003, California outbreak of END in backyard fowl and commercial poultry resulted in the destruction of about 3.3 million birds and cost \$200 million dollars to control the disease [11,12]. Outbreaks of ND have been reported in many countries with considerable economic losses [1]. Such outbreaks warrant continuous surveillance for END in commercial poultry and wild birds.

The surveillance of NDVs in waterfowl is sporadic and often occurs with other monitoring programs such as those for avian influenza viruses (AIV) [13,14]. Wild birds are considered the natural reservoirs of NDVs and mostly harbor lentogenic strains. Studies on genetic diversity among lentogenic strains of NDVs revealed that some of the NDVs from waterfowl and shorebirds were phylogenetically related with NDVs isolated from live-bird markets in the US [4]. It is recommended that epidemiological studies should be continued to determine the prevalence of lentogenic NDVs in wild bird populations [4]. An epidemiological link between isolates recovered from outbreaks in domestic poultry with those obtained from wild bird populations has also been suggested [8,9,15,16]. Therefore, continuous surveillance of wild bird populations may help better understand the NDVs circulating in the environment. This study was conducted to characterize NDV isolates obtained from waterfowl samples. In this study, the cloacal samples from waterfowl from Upper Midwest region of the US were initially screened for AIV by real time reverse transcription-polymerase chain reaction (rRT-PCR); the AIV positive samples by rRT-PCR were inoculated on to the embryonated eggs for virus isolation that yielded NDV in some of them. The NDV isolates were characterized by sequencing to determine the pathotypes and genotypes involved and the changes at the nucleotide and amino acid levels.

## Results

Altogether, 159 viral isolations from cloacal samples of AIV rRT-PCR-positive waterfowl ( $n = 890$ ) were obtained, as shown by hemagglutinating (HA) activity of allantoic fluid in embryonated eggs. Of these, 43 were positive for NDV by reverse transcription-polymerase chain reaction (RT-PCR). BLAST analysis of partial sequences of F gene of NDV isolates confirmed their identity. These isolates were obtained from 24 mallards (MALL; *Anas platyrhynchos*), seven American green-winged teals (AGWT; *Anas crecca*), six northern pintails (NOPI; *Anas acuta*), four blue-winged teals (BWTE; *Anas discors*), and two wood ducks (WODU; *Aix sponsa*). Spatial distribution revealed that 28 isolates were obtained from South Dakota, 14 from Minnesota, and 1 from North Dakota.

### Cleavage site analysis

The F gene portion (333 base pairs) corresponding to nucleotide positions 170-502 of GenBank accession number [AF217084](#) was sequenced. Deduced amino acid sequences of the F gene cleavage site were used to determine the pathotypes involved and are shown in Table 1. The fusion gene of virulent NDVs is characterized by the presence of a pair of dibasic amino acids at the cleavage site while in lentogenic strains it is characterized by the presence of monobasic amino acids. None of the isolates had the sequence motif of <sup>111</sup>GR/KRQRK/RF<sup>117</sup>, a characteristic of the virulent strains. All 43 NDVs had an avirulent motif of monobasic amino acids at their F gene cleavage sites. Of the 43 isolates, 23 exhibited sequence motif of <sup>111</sup>GGKQGRL<sup>117</sup>, 19 exhibited the sequence motif of <sup>111</sup>GEKQGRL<sup>117</sup>, and one isolate exhibited the sequence motif of <sup>111</sup>GERQGRL<sup>117</sup> at the cleavage site of F gene.

### Phylogenetic analysis

Phylogenetic analysis of partial F gene nucleotide sequences of NDV isolates was done by comparing them with already published F gene sequences of both class I and class II NDVs. None of the isolates clustered with class I NDVs (Figure 1); all isolates clustered with class II NDVs (Figure 1). Within class II, all isolates clustered with genotype I or II. Five of the 43 isolates clustered with NDV sequences of genotype I/Ia suggesting them to belong to genotype I (Figure 1). Four of the five isolates clustered together with genotype I NDVs from the US [Mallard/US(MD)/04-483/2004, [EF564942](#); Mallard/US(MD)/04-204/2004, [EF564821](#); and Mallard/US(MD)/04-235/2004, [EF564901](#)] and Korea [KR/duck/05/07, [EU547755](#)]. The sequence homology among these four isolates was 99.6% to 100% at the nucleotide level. The remaining one isolate was in a different group from these four isolates and was phylogenetically closer to genotype I NDVs from China [Heb02, [AY427817](#)], the US [AV 80/97 D813-2, [AY175736](#)] and Ireland [AV 963/98 NZ5/97,

**Table 1: Details of Newcastle disease viral isolates of this study.**

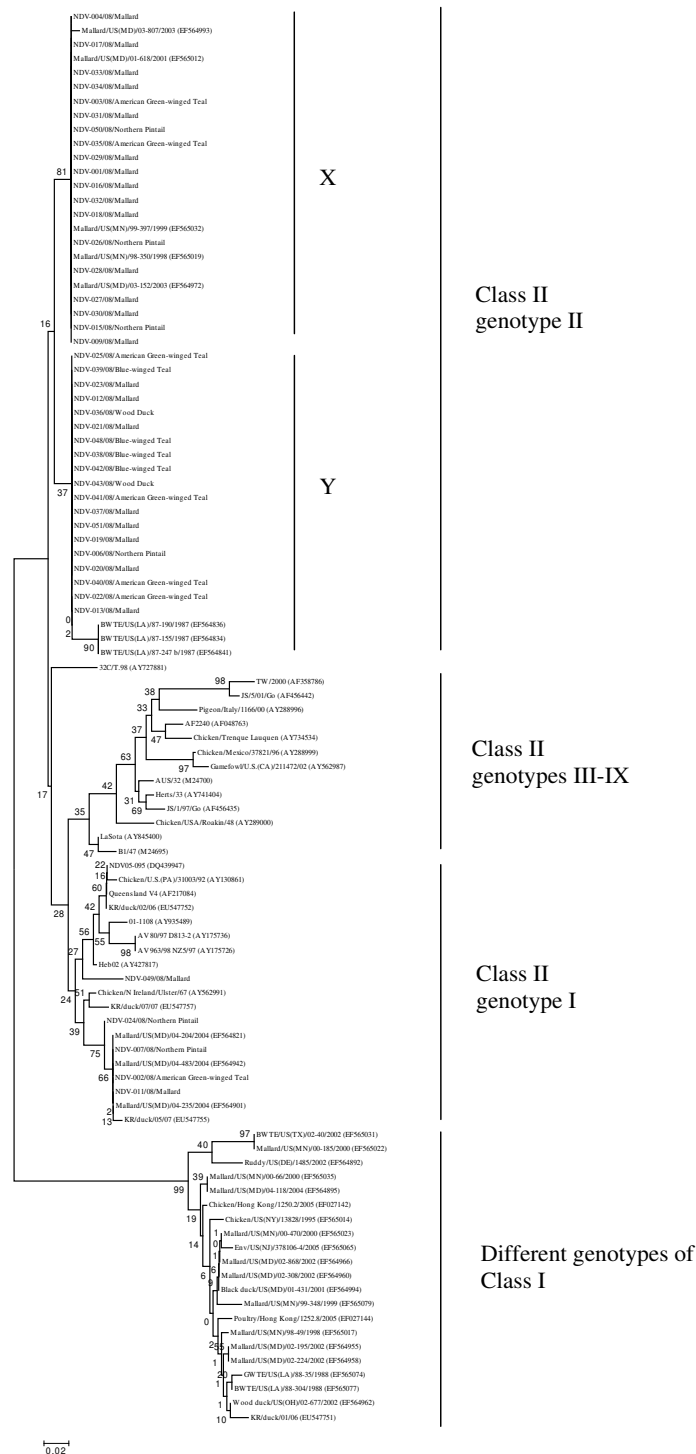
GenBank accession number	Isolate name	Fusion gene cleavage site (I11-I17)	Class	Genotype	Species	State	Country
<a href="#">GQ229531</a>	NDV-001/US(MN)/2008	GEKQGR	II	II	Mallard	Minnesota	USA
<a href="#">GQ229532</a>	NDV-002/US(MN)/2008	GGKQGR	II	I	AGWT <sup>A</sup>	Minnesota	USA
<a href="#">GQ229533</a>	NDV-003/US(MN)/2008	GEKQGR	II	II	AGWT	Minnesota	USA
<a href="#">GQ229534</a>	NDV-004/US(MN)/2008	GEKQGR	II	II	Mallard	Minnesota	USA
<a href="#">GQ229535</a>	NDV-006/US(MN)/2008	GGKQGR	II	II	Northern pintail	Minnesota	USA
<a href="#">GQ229536</a>	NDV-007/US(SD)/2008	GGKQGR	II	I	Northern pintail	South Dakota	USA
<a href="#">GQ229537</a>	NDV-009/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229538</a>	NDV-011/US(SD)/2008	GGKQGR	II	I	Mallard	South Dakota	USA
<a href="#">GQ229539</a>	NDV-012/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229540</a>	NDV-013/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229541</a>	NDV-015/US(SD)/2008	GEKQGR	II	II	Northern pintail	South Dakota	USA
<a href="#">GQ229542</a>	NDV-016/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229543</a>	NDV-017/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229544</a>	NDV-018/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229545</a>	NDV-019/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229546</a>	NDV-020/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229547</a>	NDV-021/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229548</a>	NDV-022/US(SD)/2008	GGKQGR	II	II	AGWT	South Dakota	USA
<a href="#">GQ229549</a>	NDV-023/US(SD)/2008	GGKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229550</a>	NDV-024/US(SD)/2008	GGKQGR	II	I	Northern pintail	South Dakota	USA
<a href="#">GQ229551</a>	NDV-025/US(SD)/2008	GGKQGR	II	II	AGWT	South Dakota	USA
<a href="#">GQ229552</a>	NDV-026/US(SD)/2008	GEKQGR	II	II	Northern pintail	South Dakota	USA
<a href="#">GQ229553</a>	NDV-027/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229554</a>	NDV-028/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229555</a>	NDV-029/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229556</a>	NDV-030/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229557</a>	NDV-031/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229558</a>	NDV-032/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229559</a>	NDV-033/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229560</a>	NDV-034/US(SD)/2008	GEKQGR	II	II	Mallard	South Dakota	USA
<a href="#">GQ229561</a>	NDV-035/US(SD)/2008	GEKQGR	II	II	AGWT	South Dakota	USA
<a href="#">GQ229562</a>	NDV-036/US(MN)/2008	GGKQGR	II	II	Wood duck	Minnesota	USA
<a href="#">GQ229563</a>	NDV-037/US(MN)/2008	GGKQGR	II	II	Mallard	Minnesota	USA
<a href="#">GQ229564</a>	NDV-038/US(MN)/2008	GGKQGR	II	II	Blue-winged teal	Minnesota	USA
<a href="#">GQ229565</a>	NDV-039/US(MN)/2008	GGKQGR	II	II	Blue-winged teal	Minnesota	USA
<a href="#">GQ229566</a>	NDV-040/US(MN)/2008	GGKQGR	II	II	AGWT	Minnesota	USA
<a href="#">GQ229567</a>	NDV-041/US(MN)/2008	GGKQGR	II	II	AGWT	Minnesota	USA
<a href="#">GQ229568</a>	NDV-042/US(MN)/2008	GGKQGR	II	II	Blue-winged teal	Minnesota	USA
<a href="#">GQ229569</a>	NDV-043/US(MN)/2008	GGKQGR	II	II	Wood duck	Minnesota	USA
<a href="#">GQ229570</a>	NDV-048/US(SD)/2008	GGKQGR	II	II	Blue-winged teal	South Dakota	USA
<a href="#">GQ229571</a>	NDV-049 US(MN)/2008	GERQGR	II	I	Mallard	Minnesota	USA
<a href="#">GQ229572</a>	NDV-050/US(SD)/2008	GEKQGR	II	II	Northern pintail	South Dakota	USA
<a href="#">GQ229573</a>	NDV-051/US(ND)/2008	GGKQGR	II	II	Mallard	North Dakota	USA

<sup>A</sup>AGWT = American Green-winged teal

[AY175726](#)]. This isolate had sequence homology of 90.9% to 90.4% at nucleotide level with the other four isolates of genotype I of this study. All five genotype I isolates had sequence homology of 87.9% to 100% with class II genotype I NDVs used for comparison.

The remaining 38 isolates clustered with genotype II NDVs. These isolates clustered into two groups with 19 isolates in each group. For ease of understanding, we have named these two groups as X and Y (Figure 1). The isolates

in group X were phylogenetically close to genotype IIa NDVs from wild birds from different regions of the US [Mallard/US(MD)/03-152/2003, [EF564972](#); Mallard/US(MD)/01-618/2001, [EF565012](#); Mallard/US(MN)/99-397/1999, [EF565032](#); Mallard/US(MN)/98-350/1998, [EF565019](#); and Mallard/US(MD)/03-807/2003, [EF564993](#)]. The isolates in group X were also phylogenetically close to a genotype IIa NDV from Argentina [32C/T.98, [AY727881](#)], but the latter was in a different group. None of the already reported NDV sequences of class II



**Figure 1**  
**Phylogenetic tree based on partial nucleotide sequences [corresponding to nucleotide positions 170-502 of GenBank: AF217084] of fusion gene of Newcastle disease virus.** The sequences starting with NDV (without accession numbers) are from the present study, and the sequences with virus name (GenBank accession numbers) are previously published sequences of NDVs. The phylogenetic tree was constructed by Neighbor-Joining method, 500 bootstrap replicates (bootstrap values are shown on tree).

genotype II used for comparison clustered together with NDV isolates of group Y. Though the isolates in group Y were phylogenetically close to already reported genotype IIa NDVs from wild birds in the US [Blue winged teal/US(LA)/87-190/1987, [EF564836](#); Blue winged teal/US(LA)/87-155/1987, [EF564834](#); Blue winged teal/US(LA)/87-247\_b/1987, [EF564841](#)], they were not in the same group. The vaccine strains [LaSota, [AY845400](#); B1, [M24695](#)] clustered in a different group from isolates of this study. All already published sequences of velogenic strains with in class II were phylogenetically distinct from NDVs of this study (Figure 1). The sequence homology of genotype II isolates of this study ranged from 95.5% to 100% at the nucleotide level, and the homology as compared to already published sequences of class II genotype II ranged from 90.4% to 100%.

## Discussion

This study was conducted to characterize NDVs isolated from waterfowl in the Upper Midwest region of the US. The initial aim of this study was to isolate and characterize AIV from waterfowl. During the study period, 7458 cloacal samples were collected and of these, 11.9% samples were AIV positive by rRT-PCR. Inoculation of these AIV positive samples in embryonated chicken eggs yielded hemagglutinating viruses and of these, 43 were identified as NDVs by RT-PCR using primer specific for F gene. We were expecting the isolation of AIV rather than NDV on inoculation in embryonated eggs as the samples were initially positive for AIV by rRT-PCR. The possibility of the presence of other hemagglutinating virus(es) in HA positive-AIV negative (by RT-PCR for matrix gene)-NDV negative (by RT-PCR for F gene) allantoic fluid cannot be ruled out and testing of such allantoic fluid is underway in our laboratory. The isolation of NDV from samples that were rRT-PCR positive for AIV indicates that the cloacal sample may have mixed infection with NDV and AIV with concentration of NDV being higher than that of AIV. Hence, the NDV probably overgrew AIV upon inoculation in embryonated chicken eggs. It is to be noted that we tested only AI rRT-PCR positive samples by inoculation in embryonated eggs; testing of more samples might have led to isolation of more NDVs. The isolation of NDV from AIV positive samples indicates the presence of both viruses (AIV and NDV) in waterfowl. The AIV positive allantoic fluid by RT-PCR was not tested for NDV; this testing might provide a better picture of mixed infection of both NDV and AIV. Mixed infection of AIV and NDV in waterfowl has been reported earlier [17,18].

A large amount of sequence data on NDVs isolated throughout the world has been published over the years and is now available for sequence comparison and phylogenetic analysis which can be used to predict the pathotypes and to determine the origin of NDV outbreaks. It has

been well established that cleavage of NDV fusion protein is a major determinant for viral virulence. In this study, the F gene sequence of NDVs was used for pathotyping as well as their characterization into different classes and genotypes. None of the isolates was found to be velogenic on the basis of sequence motif of F gene cleavage site. It has been reported that virulent virus has at least one pair of basic amino acids at residues 115 and 116 plus a phenylalanine at residue 117 and a basic amino acid (R) at 113 at the cleavage site whereas lentogenic strains lack dibasic amino acids [19]. All NDV isolates of this study had lentogenic motif at the cleavage site. These results are in agreement with previous studies reporting the detection of lentogenic NDVs in wild birds and domestic ducks [4,9,15,20,21]. None of the isolates had the sequence motif of <sup>111</sup>GERQE/DRL<sup>117</sup> of class I isolates, although the latter have been reported in wild birds and domestic ducks [4,21]. For example, [4] reported seven of the nine genotypes of class I NDVs in waterfowl and shore birds in the US while [21] reported the presence of class I genotype 2 NDVs in domestic ducks in Korea.

Of the 43 isolates, 42 had the sequence motif of <sup>111</sup>GG/EKQGRL<sup>117</sup> at the cleavage site and were phylogenetically similar to either genotype I or genotype II within class II. This sequence motif has been reported earlier in genotypes I and II of class II NDVs [4]. However, a different sequence motif (<sup>111</sup>GRRQRRF<sup>117</sup>) was reported in the lentogenic strains from Australia [22]. One of the isolates had the sequence motif of <sup>111</sup>GERQGRL<sup>117</sup> and this isolate also clustered with class II genotype I strains. This isolate differed from other 42 isolates in the sense that the amino acid lysine was replaced by arginine at position 113.

Overall genotype II viruses were more predominant than genotype I viruses in this study. This finding has the support of [4] who also observed more genotype IIa viruses than genotype I viruses within class II. The NDV isolates in this study were derived only from rRT-PCR AIV positive samples, the possibility of presence of genotypes of both classes (that were not detected in this study) in rRT-PCR AIV negative samples cannot be ruled out. Within class II, the NDV sequences clustered into two different groups. None of the isolates was phylogenetically close to vaccine strains used for comparison. This indicates that in spite of the regular use of live vaccines in poultry throughout the world, their transmission to wild birds may not be a common phenomenon. In an earlier study, [4] also did not detect any vaccine strains in wild birds in the US. Since wild birds have been reported to be a reservoir of NDV [16,23], the mixing of different species at stop-overs during migration and the sharing of common wintering and breeding areas may provide opportunity for virus spread within and between countries and may help perpetuate different genotypes and classes of NDVs in these birds.

The phylogenetic proximity of our isolates with those from the US, China, Korea, and Ireland points to this likelihood.

The presence of class II viruses in wild birds is of concern because this class of viruses has been responsible for several panzootics of Newcastle disease in poultry [24,25]. There are reports suggesting that velogenic NDVs might arise from lentogenic NDVs in nature [23,26]. Further, studies have also suggested that point mutation, and not gene recombination, may be responsible for generating virulent and avirulent strains. For example, the NDV outbreak in Australian poultry during 1998-2000 was caused by a virulent NDV that originated due to mutation in a class II genotype I virus [26]. These authors were of the opinion that lentogenic viruses have the potential to become virulent with the passage of time. Even passaging of NDVs from one host to another has been reported to increase their virulence [16,27]. In addition, the selective forces imposed by a new host environment may also play a role in acquisition of virulence [28]. These findings suggest that the lentogenic strains from wild birds may acquire virulence by waterfowl-to-domestic poultry transmission in nature. In such a scenario we may encounter an NDV outbreak in domestic poultry.

Similar to low pathogenic AIV, the lentogenic NDVs in wild bird populations invariably do not cause obvious disease. Even virulent strains of NDVs that are lethal to chickens, have been isolated from apparently healthy

domestic ducks [14,29,30]. Though virulent strains of NDVs were not detected in this study, their presence in the population cannot be ruled out in view of the potential created by the comingling nature and migration patterns of wild birds within and across continents. Thus, continuous surveillance for NDV in wild birds is essential for better understanding of its epidemiology. In conclusion, the present study reveals the circulation of class II (genotypes I and II) lentogenic strains of NDVs in wild birds in the Upper Midwest region of the US. Further studies are needed to determine the true prevalence and implications of various genotypes of NDV within wild bird population.

## Conclusion

This study indicates the circulation of class II genotypes I and II NDVs in waterfowl in the Upper Midwest region of the US with an avirulent motif of monobasic amino acids at their F gene cleavage sites. Phylogenetically distant relationship of NDVs of this study with vaccine strains indicates that in spite of the regular use of live vaccines in poultry, their transmission to wild birds may not be a common phenomenon.

## Methods

### Sample collection

Under an NIH funded surveillance program on avian influenza, cloacal and oropharyngeal (OP) swabs were collected from various waterfowl species in Minnesota, South Dakota, and North Dakota from April 2008 to October 2008. The swabs were placed in brain heart infu-

**Table 2: Previously published F gene sequences of class I Newcastle disease virus used for phylogenetic analysis.**

GenBank accession number	Strain name	Fusion cleavage site	Genotype	Country
<a href="#">EF565077</a>	Blue winged teal/US(LA)/88-304/1988	GERQERL	1	USA
<a href="#">EF565074</a>	Green winged teal/US(LA)/88-35/1988	GERQERL	1	USA
<a href="#">EF565014</a>	Chicken/US(NY)/13828/1995	GERQERL	1	USA
<a href="#">EF564958</a>	Mallard/US(MD)/02-224/2002	GERQERL	2	USA
<a href="#">EF565017</a>	Mallard/US(MN)/98-49/1998	GERQERL	2	USA
<a href="#">EF564962</a>	Wood duck/US(OH)/02-677/2002	GERQERL	2	USA
<a href="#">EF564955</a>	Mallard/US(MD)/02-195/2002	GERQERL	2	USA
<a href="#">EU547751</a>	KR/duck/01/06	GERQERL	2	Korea
<a href="#">EF027144</a>	Poultry/Hong Kong/1252.8/2005	GERQERL	3	Hong Kong
<a href="#">EF027142</a>	Chicken/Hong Kong/1250.2/2005	GERQERL	3	Hong Kong
<a href="#">EF564960</a>	Mallard/US(MD)/02-308/2002	GERQERL	4	USA
<a href="#">EF564966</a>	Mallard/US(MD)/02-868/2002	GERQERL	4	USA
<a href="#">EF564994</a>	Black duck/US(MD)/01-431/2001	GERQERL	5	USA
<a href="#">EF565023</a>	Mallard/US(MN)/00-470/2000	GERQERL	5	USA
<a href="#">EF565079</a>	Mallard/US(MN)/99-348/1999	GERQERL	5	USA
<a href="#">EF565065</a>	Environment/US(NJ)/378106-4/2005	GERQERL	6	USA
<a href="#">EF565035</a>	Mallard/US(MN)/00-66/2000	GERQERL	7	USA
<a href="#">EF564895</a>	Mallard/US(MD)/04-118/2004	GERQERL	7	USA
<a href="#">EF564892</a>	Ruddy turnstone/US(DE)/1485/2002	GERQERL	8	USA
<a href="#">EF565031</a>	Blue winged teal/US(TX)/02-40/2002	GERQERL	9	USA
<a href="#">EF565022</a>	Mallard/US(MN)/00-185/2000	GERQERL	9	USA

**Table 3: Previously published F gene sequences of class II Newcastle disease virus used for phylogenetic analysis.**

GenBank accession number	Strain name	Fusion cleavage site	Genotype	Country
<a href="#">AY175726</a>	AV 963/98 NZ5/97 (GNZDK98025)	GGKQGRL	I	Ireland
<a href="#">AY562991</a>	Chicken/N. Ireland/Ulster/67	GGKQGRL	I	Ireland
<a href="#">DQ439947</a>	NDV05-095	GGKQGRL	I	China
<a href="#">EF564821</a>	Mallard/US(MD)/04-204/2004	GGKQGRL	I	USA
<a href="#">EF564901</a>	Mallard/US(MD)/04-235/2004	GGKQGRL	I	USA
<a href="#">EF564942</a>	Mallard/US(MD)/04-483/2004	GGKQGRL	I	USA
<a href="#">AY130861</a>	Chicken/U.S.(PA)/31003/92	GGKQGRL	I	USA
<a href="#">AF217084</a>	Queensland V4	GGKQGRL	I	Australia
<a href="#">AY175736</a>	AV 80/97 D813-2 (HTWDK95193)	GGKQGRL	Ia	USA
<a href="#">AY427817</a>	Heb02	GGKQGRL	Ia	China
<a href="#">AY935489</a>	01-1108	GRRQGRL	Ia	Australia
<a href="#">AY289000</a>	Chicken/USA/Roakin/48	GRRQKRF	II	USA
<a href="#">AY845400</a>	LaSota	GGRQGRL	II	China
<a href="#">M24695</a>	BI/47	GGRQGRL	II	USA
<a href="#">AY727881</a>	32C/T.98	GGKQGRL	Ila	Argentina
<a href="#">EF564834</a>	Blue winged teal/US(LA)/87-155/1987	GGKQGRL	Ila	USA
<a href="#">EF564836</a>	Blue winged teal/US(LA)/87-190/1987	GGKQGRL	Ila	USA
<a href="#">EF564841</a>	Blue winged teal/US(LA)/87-247_b/1987	GGKQGRL	Ila	USA
<a href="#">EF564972</a>	Mallard/US(MD)/03-152/2003	GEKQGRL	Ila	USA
<a href="#">EF564993</a>	Mallard/US(MD)/03-807/2003	GEKQGRL	Ila	USA
<a href="#">EF565012</a>	Mallard/US(MD)/01-618/2001	GEKQGRL	Ila	USA
<a href="#">EF565019</a>	Mallard/US(MN)/98-350/1998	GEKQGRL	Ila	USA
<a href="#">EF565032</a>	Mallard/US(MN)/99-397/1999	GEKQGRL	Ila	USA
<a href="#">EU547752</a>	KR/duck/02/06	GGKQGRL	II	Korea
<a href="#">EU547755</a>	KR/duck/05/07	GGKQGRL	II	Korea
<a href="#">EU547757</a>	KR/duck/07/07	GGKQGRL	II	Korea
<a href="#">M24700</a>	AUS/32	GRRQKRF	III	Australia
<a href="#">AY741404</a>	Herts/33	GRRQRRF	IV	USA
<a href="#">AY288999</a>	Chicken/Mexico/37821/96	GRRQKRF	V	Mexico
<a href="#">AY562987</a>	Gamefowl/U.S.(CA)/211472/02	GRRQKRF	V	USA
<a href="#">AY288996</a>	Pigeon/Italy/1166/00	GRRQKRF	VI	Italy
<a href="#">AF358786</a>	TW/2000	GRRQKRF	VII	Taiwan
<a href="#">AF456442</a>	JS/5/01/Go	GRRQKRF	VII	China
<a href="#">AF048763</a>	AF2240	GRRQKRF	VIII	Malaysia
<a href="#">AY734534</a>	Chicken/Trenque Lauquen	GRRQKRF	VIII	Argentina
<a href="#">AF456435</a>	JS/1/97/Go	GRRQKRF	IX	China

sion broth containing antibiotics (penicillin 500 IU/mL, streptomycin 500 µg/mL, neomycin 0.15 mg/mL, fungizone 1.5 µg/mL, and gentamicin 50 µg/mL) and were transported on ice to the laboratory. The initial aim of the project was to test cloacal samples (n = 7458) from waterfowl species for the detection of AIV for which five samples each were pooled and the pools were tested for AIV using rRT-PCR [31]. Individual samples in positive pools were then tested for the detection of AIV by rRT-PCR.

#### Virus isolation

Individual samples positive for AIV by rRT-PCR (n = 890) were inoculated in 9-day-old specific pathogen free embryonated chicken eggs for virus isolation (VI). Allantoic fluid from inoculated eggs was harvested four days post inoculation and subsequently tested for hemagglutination (HA) using 0.5% turkey erythrocytes. The HA pos-

itive allantoic fluids (n = 159) were tested by RT-PCR for the confirmation of AIV as described below.

#### Total RNA extraction and RT-PCR

Total RNA was extracted from allantoic fluids and a known AIV isolate using QIAamp Viral RNA extraction kit (Qiagen, Valencia, CA). Extracted RNAs were subjected to RT-PCR using primers targeting the matrix gene of AIV [32]. A band of 1027 base pairs was observed in 52 cases indicating them to be AIV. The HA positive allantoic fluids that were negative for AIV (n = 107) were then tested for NDV by RT-PCR. Total RNA extracted from a known APMV-1 was used as a positive control. The RNA was amplified using primers specific to the F gene of NDV [33]. PCR amplification was carried out using Qiagen OneStep RT-PCR kit (Qiagen, Valencia, CA). Amplified PCR products were electrophoresed on 1.2% agarose gel.

A band of 356 base pairs was observed in 43 cases indicating them to be NDVs. Further studies are underway to determine the identity of the remaining HA positive allantoic fluids (n = 64). The NDV positive PCR products were purified using a PCR purification kit (Qiagen, Valencia, CA) and were then sequenced in both directions at the BioMedical Genomic Center, University of Minnesota.

### Phylogenetic analysis

The forward and reverse nucleotide sequences of all 43 isolates were curated, edited and aligned using a "Sequencher" software <http://www.msi.umn.edu>. The aligned sequences were analyzed on NCBI website <http://www.ncbi.nlm.nih.gov> using BLAST to confirm their identity. The nucleotide sequences were then aligned using MEGA 4.0 software by Clustal W method. The evolutionary distances were computed by Pairwise Distance method using the Maximum Composite Likelihood Model. A phylogenetic tree of aligned sequences was constructed by Neighbor-Joining method (500 replicates for bootstrap). The F gene nucleotide sequences [corresponding to nucleotide positions 170-502 of GenBank: [AF217084](#)] were translated to deduced amino acid sequences to determine the pathotype involved. The nucleotide sequences were also compared with NDV sequences available in the GenBank. The virus types and their GenBank accession numbers used for comparison are given in Tables 2 and 3. These included F gene sequences of different genotypes of class I and class II NDVs. To maintain uniformity and consistency, class I genotypes are indicated using Arabic numerals (1-9) while class II genotypes are indicated using Roman numerals (I-IX).

### GenBank accession numbers

The NDV sequence data were submitted to the GenBank database; the accession numbers and other details are shown in Table 1.

### Abbreviations

AGWT: American green-winged teal; AIV: avian influenza virus; APMV: avian paramyxovirus; END: exotic Newcastle disease; HA: hemagglutination; MALL: mallard; ND: Newcastle disease; NDV: Newcastle disease virus; NOPI: northern pintail; rRT-PCR: real time reverse-transcription polymerase chain reaction; RT-PCR: reverse-transcription polymerase chain reaction; VI: virus isolation; WODU: wood duck.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

NJ and YC contributed for RT-PCR, sequence analysis and generation of phylogenetic tree. MA and AKC performed

the virus isolation in eggs. NJ and SMG drafted the manuscript. SMG coordinated overall planning and designed this study. PTR coordinated sample collection from wild birds from Minnesota, South Dakota, and North Dakota. All authors' have read and approved the final manuscript.

### Acknowledgements

This work has been funded in whole or in part with federal funds from the National Institute of Allergy and Infectious Diseases, National Institute of Health, Department of Health and Human Services, under Contract No. HHSN266200700007C. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the NIH.

### References

- Alexander DJ: **Newcastle disease, other avian paramyxoviruses and pneumovirus infections.** In *Diseases of Poultry* 11th edition. Edited by: Saif YM, Barnes HJ, Glisson JR, Fadly AM, McDougald LR, Swayne DE. Ames, IA: Iowa State University Press; 2003.
- Seal BS, King DJ, Bennett JD: **Characterization of Newcastle disease virus isolates by reverse transcription PCR coupled to direct nucleotide sequencing and development of sequence database for pathotype prediction and molecular epidemiological analysis.** *J Clin Microbiol* 1995, **33**:2624-2630.
- Czegledi A, Ujvari D, Somogyi E, Wehmann E, Werner O, Lomniczi B: **Third genome size category of avian paramyxovirus serotype 1 (Newcastle disease virus) and evolutionary implications.** *Virus Res* 2006, **120**:36-48.
- Kim LM, King DJ, Curry PE, Suarez DL, Swayne DE, Stallknecht DE, Slemmons RD, Pedersen JC, Senne DA, Winker K, Afonso CL: **Phylogenetic diversity among low-virulence Newcastle disease viruses from waterfowl and shorebirds and comparison of genotype distributions to those of poultry-origin isolates.** *J Virol* 2007, **81**:12641-12653.
- Liu XF, Wan HQ, Ni XX, Wu YT, Liu WB: **Pathotypical and genotypical characterization of strains of Newcastle disease virus isolated from outbreaks in chicken and goose flocks in some regions of China during 1985-2001.** *Arch Virol* 2003, **148**:1387-1403.
- Marin MC, Villegas P, Bennett JD, Seal BS: **Virus characterization and sequence of the fusion protein gene cleavage site of recent Newcastle disease virus field isolates from the southeastern United States and Puerto Rico.** *Avian Dis* 1996, **40**:382-390.
- King DJ, Seal BS: **Biological and molecular characterization of Newcastle disease virus isolates from surveillance of live bird markets in the northeastern United States.** *Avian Dis* 1997, **41**:683-689.
- Jorgensen PH, Handberg KJ, Ahrens P, Therkildsen OR, Manvell RJ, Alexander DJ: **Strains of avian paramyxovirus type 1 of low pathogenicity for chickens isolated from poultry and wild birds in Denmark.** *Vet Rec* 2004, **154**:497-500.
- Huovilainen A, Ek-Kommonen C, Manvell R, Kinnunen L: **Phylogenetic analysis of avian paramyxovirus 1 strains isolated in Finland.** *Arch Virol* 2001, **146**:1775-1785.
- Seal BS, King DJ, Locke DP, Senne DA, Jackwood MW: **Phylogenetic relationship among highly virulent Newcastle disease virus isolates obtained from exotic birds and poultry from 1989 to 1996.** *J Clin Microbiol* 1998, **36**:1141-1145.
- Pedersen JC, Senne DA, Woolcock PR, Kinde H, King DJ, Wise MG, Panigrahy B, Seal BS: **Phylogenetic relationships among virulent Newcastle disease virus isolates from the 2002-2003 outbreak in California and other recent outbreaks in North America.** *J Clin Microbiol* 2004, **42**:2329-2334.
- Wise MG, Suarez DL, Seal BS, Pedersen JC, Senne DA, King DJ, Kapczynski DR, Spackman E: **Development of a real-time reverse-transcription PCR for detection of Newcastle disease virus RNA in clinical samples.** *J Clin Microbiol* 2004, **42**:329-338.
- Alfonso CP, Cowen BS, van Campen H: **Influenza A viruses isolated from waterfowl in two wildlife management areas of Pennsylvania.** *J Wildlife Dis* 1995, **31**:179-185.



14. Hanson BA, Swayne DE, Senne DA, Lobpries DS, Hurst J, Stallknecht DE: **Avian influenza viruses and paramyxoviruses in wintering and resident ducks in Texas.** *J Wildlife Dis* 2005, **41**:624-628.
15. Takakuwa H, Ito T, Takada A, Okazaki K, Kida H: **Potentially virulent Newcastle disease viruses are maintained in migratory waterfowl populations.** *Jpn J Vet Res* 1998, **45**:207-215.
16. Shengqing Y, Kishida N, Ito H, Kida H, Otsuki K, Kawaoka Y, Ito T: **Generation of velogenic Newcastle disease viruses from a nonpathogenic waterfowl isolate by passaging in chickens.** *Virology* 2002, **301**:206-211.
17. Lindh E, Huovilainen A, Ratti O, Ek-Kommonen C, Sironen T, Huhtamo E, Poysa H, Vaeheri A, Vapalahti O: **Orthomyxo-, paramyxo- and flavivirus infections in wild waterfowl in Finland.** *Virol J* 2008, **5**:35.
18. Jahangir A, Ruenphet S, Ueda S, Ueno Y, Shoham D, Shindo J, Okamura M, Nakamura M, Takehara K: **Avian influenza and Newcastle disease viruses from northern pintail in Japan: Isolation, characterization and inter-annual comparisons during 2006-2008.** *Virus Res* 2009, **143**:44-52.
19. The World Organization for Animal Health (OIE): *Manual of diagnostic tests and vaccines for terrestrial animals, Newcastle disease. Part 2, Section 2.3, Chapter 2.3.14* 2008 [[http://www.oie.int/eng/normes/mmanual/A\\_INDEXTM](http://www.oie.int/eng/normes/mmanual/A_INDEXTM)]. Paris: OIE
20. Sakai K, Sakabe G, Tani O, Watanabe Y, Jahangir A, Nakamura M, Takehara K: **Characterization of Newcastle disease virus isolated from Northern Pintail (*Anas acuta*) in Japan.** *J Vet Med Sci* 2007, **69**:1307-1311.
21. Lee EK, Jeon WJ, Kwon JH, Yang CB, Choi KS: **Molecular epidemiological investigation of Newcastle disease virus from domestic ducks in Korea.** *Vet Microbiol* 2009, **134**:241-248.
22. Westbury H: **Commentary. Newcastle disease virus: an evolving pathogen.** *Avian Pathol* 2001, **30**:5-11.
23. Collins MS, Bashiruddin JB, Alexander DJ: **Deduced amino acid sequences at the fusion protein cleavage site of Newcastle disease viruses showing variation in antigenicity and pathogenicity.** *Arch Virol* 1993, **128**:363-370.
24. Alexander DJ: *Newcastle disease* Boston: Kluwer Academic Publishers; 1988.
25. Ballagi-Pordany A, Wehmann E, Herczeg J, Belak S, Lomniczi B: **Identification and grouping of Newcastle disease virus strains by restriction site analysis of a region from the F gene.** *Arch Virol* 1996, **141**:243-261.
26. Gould AR, Kattenbelt JA, Selleck P, Hansson E, Della-Porta A, Westbury HA: **Virulent Newcastle disease in Australia: Molecular epidemiological analysis of viruses isolated prior to and during the outbreaks of 1998-2000.** *Virus Res* 2001, **77**:51-60.
27. Alexander DJ, Parsons G: **Protection of chickens against challenge with the variant virus responsible for Newcastle disease in 1984 by conventional vaccination.** *Vet Rec* 1986, **118**:176-177.
28. Zanetti F, Berinsein A, Carrillo E: **Effect of host selective pressure on Newcastle disease virus virulence.** *Microbial Pathog* 2008, **44**:135-140.
29. Tsai HJ, Chang KH, Tseng CH, Frost KM, Manvell RJ, Alexander DJ: **Antigenic and genotypical characterization of Newcastle disease viruses isolated in Taiwan between 1969 and 1996.** *Vet Microbiol* 2004, **104**:19-30.
30. Liu H, Wang Z, Wu Y, Zheng D, Sun C, Bi D, Zuo Y, Xu T: **Molecular epidemiological analysis of Newcastle disease virus isolated in China in 2005.** *J Virol Methods* 2007, **140**:206-211.
31. Spackman E, Senne DA, Myers TJ, Bulaga LL, Garber LP, Perdue ML, Lohman K, Daum LT, Suarez DL: **Development of a real-time reverse transcriptase PCR assay for type A influenza virus and the avian H5 and H7 hemagglutinin subtypes.** *J Clin Microbiol* 2002, **40**:3256-3260.
32. Chan CH, Lin KL, Chan Y, Wang YL, Chi YT, Tu HL, Shieh HK, Liu WT: **Amplification of the entire genome of influenza A virus H1N1 and H3N2 subtypes by reverse-transcription polymerase chain reaction.** *J Virol Methods* 2006, **136**:38-43.
33. Nanthakumar T, Kataria RS, Tiwari AK, Butchiah G, Kataria JM: **Pathotyping of Newcastle disease viruses by RT-PCR and restriction enzyme analysis.** *Vet Res Commun* 2000, **24**:275-286.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

